RESOLUTION OIV-OENO 461-2012

METHODS FOR THE DETERMINATION OF NATAMYCIN IN WINE

THE GENERAL ASSEMBLY,

CONSIDERING Article 2 paragraph 2 iv of the Agreement establishing the International Organisation of Vine and Wine,

UPON THE PROPOSAL of the Sub-commission of Methods of Analysis of Must and Wine,

CONSIDERING that the OIV's activities shall to help to contribute to food safety;

RECOGNISES that specific official methods for the determination of natamycin in wine are not prescribed and methods already available can be applied.

DECIDES: to adopt the two methods of type IV below for the determination of natamycin in wine:

1. INTRODUCTION

Different methods for the determination of natamycin are used based mainly on HPLC in combination with DAD or MS detection. Estimation of the performance limits - limit of detection and quantification - relies on the responsibility of the laboratories according to accreditation systems (e.g. ISO/EN 17025/2005) employing the recommendations of the OIV (OENO 7/2000, E-AS1-10-LIMDET) or other normative guidelines.

As there is lack of a reliable interlaboratory estimate of the critical level, a decision limit of 5 µg/l is temporarily adopted until a reliable interlaboratory estimate or other robust indicators of the critical level can be obtained.

2. METHODS

2.1. Determination of natamycin (pimaricin) in wine by liquid chromatography coupled to high resolution mass spectrometry

2.1.1. SCOPE

This method describes an analytical procedure for the determination of natamycin (pimaricin) in wine. The level of natamycin is expressed in micrograms per litre (µg/l) of wine. In-house validation has been carried out using solvent solutions, red wine and

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white wine over the concentration range 5 – 2600 µg/l.

2.1.2. PRINCIPLE

The level of natamycin in wine is determined by direct injection of the sample into a liquid chromatograph with a high-resolution mass-spectrometric detection system (LC-HR/MS). Quantification is achieved using the standard addition method. The sample is initially analysed to gain an estimated concentration of natamycin. The analysis is then repeated with standard addition calibration standards more suited to the concentration of natamycin in the sample.

2.1.3. REAGENTS

2.1.3.1. Analytes

2.1.3.1.1. Natamycin (Pimaricin) > 95%

2.1.3.2. Chemicals

2.1.3.2.1. Methanol, HPLC Fluorescence grade (CAS no. 67-56-1).

2.1.3.2.2. Purified water for laboratory use, for example of EN ISO 3696 grade (water for analytical laboratory use - specification and test methods [ISO 3696:1987]).

2.1.3.2.3. Acetic acid, 100%, (CAS no. 64-19-7)

2.1.3.3. Solutions

2.1.3.3.1. Stock solution of natamycin (1000 µg/ml)

Weigh to the nearest 0.1 mg approximately 10 mg of natamycin (2.1.3.1.1) in a 10 ml amber volumetric flask and make up to the mark with methanol:water:acetic acid (2.1.3.3.4). Cap and sonicate. Calculate the actual concentration in micrograms of natamycin per millilitre of solution.

2.1.3.3.2. Working solution 1: natamycin (10 µg/ml)

Pipette 100 µl of stock solution (2.1.3.3.1) into a amber 10 ml volumetric flask and make up to the mark with methanol:water:acetic acid (2.1.3.3.4)

2.1.3.3.3. Working solution 2: natamycin (0.5 µg/ml)

Pipette 500 µl of working solution one (2.1.3.3.2) into an amber 10 ml volumetric flask and make up to the mark with methanol:water:acetic acid (2.1.3.3.4)

2.1.3.3.4. Solution of methanol:water:acetic acid (50:47:3, v/v)

Using a measuring cylinder, measure 500 ml of methanol (3.2.1) into a 1 L volumetric flask. Add 470 ml water (2.1.3.2.2) and shake to mix. Add 30 ml acetic acid (2.1.3.2.3) and shake well.

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2.1.3.3.5. Methanol, 3% acetic acid

Using a measuring cylinder add 30 ml of acetic acid (2.1.3.2.3) to a 1 L volumetric flask. Make up to the mark with methanol (2.1.3.2.1) and shake well.

2.1.3.3.6. Water, 3% acetic acid

Using a measuring cylinder add 30 ml of acetic acid (2.1.3.2.3) to a 1 L volumetric flask. Make up to the mark with water (2.1.3.2.2) and shake well.

2.1.4. APPARATUS

NOTE: An instrument or item of apparatus is listed only where it is specialised or made to a particular specification, the usual laboratory glassware and equipment being assumed to be available.

2.1.4.1. Liquid Chromatograph (LC)

Equipped with an automatic injector, a 100 µl injection loop and a high resolution mass spectrometer.

2.1.4.1.1. LC column

Capable of obtaining reproducible natamycin peaks and capable of separating the natamycin peaks from interfering peaks originating from the sample matrix and/or the solvents used.

NOTE: Depending on the type of equipment used for the analysis, the appropriate operating conditions should be optimised.

2.1.4.1.2. HPLC analysis

The following column and parameters have been found to be suitable:

Column: Waters Sunfire C18, 150 x 2.1 mm, 3.5 µm

Column temperature: 30 oC

Flow rate: 0.25 ml/min

Injection volume: 20 µl

Mobile phase A: Water:acetic acid, 97:3 (v/v) (2.1.3.3.6)

Mobile phase B: Methanol:acetic acid, 97:3 (v/v) (2.1.3.3.5)

Run time: 30 min

Autosampler tray: 8 oC

Gradient:

Time (min)	Mobile phase A (%)	Mobile phase B (%)

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0	90	10
25	10	90
27	10	90
27.1	90	10
30	90	10

2.1.4.2. Mass spectrometric detection (LC-HR/MS)

Ionisation mode: positive electrospray
Mass resolution: m/z/Δm/z
AGC target: High dynamic range
Max Inj time: 50 ms
Scan range: m/z 480-670
Sweep gas: 60 L/min
Aux gas: 5 L/min
Spray voltage: 3.75 V
Natamycin: m/z 666.31069 [M+H]+. confirmation ion m/z 503.22672
Retention time: 16.5 mins

2.1.5. EXPERIMENTAL PROCEDURE

Samples should be shaken to ensure homogeneity prior to sub-sampling.

2.1.5.1. Screening

For each wine pipette 2 ml of sample into two 2 ml Eppendorf centrifuge vials. Add 0 µl, 20 µl and of natamycin working solution 2 at 0.5 µg/ml (2.1.3.3.3) to the vials respectively. This is equivalent to 0 µg/l and 5 µg/l natamycin added. Shake the vials for one minute and then centrifuge for 10 min at 14000 rpm. Filter an aliquot through 0.2 µm PTFE into an amber 2 ml vial. Analyse by LC-HR/MS (section 6) and estimate the concentration of natamycin in the sample (section 7).

If the estimated concentration of natamycin is less than 5 µg/l report the data as < 5 µg/l. If the estimated concentration of natamycin is greater than 5 µg/l follow section 5.2.

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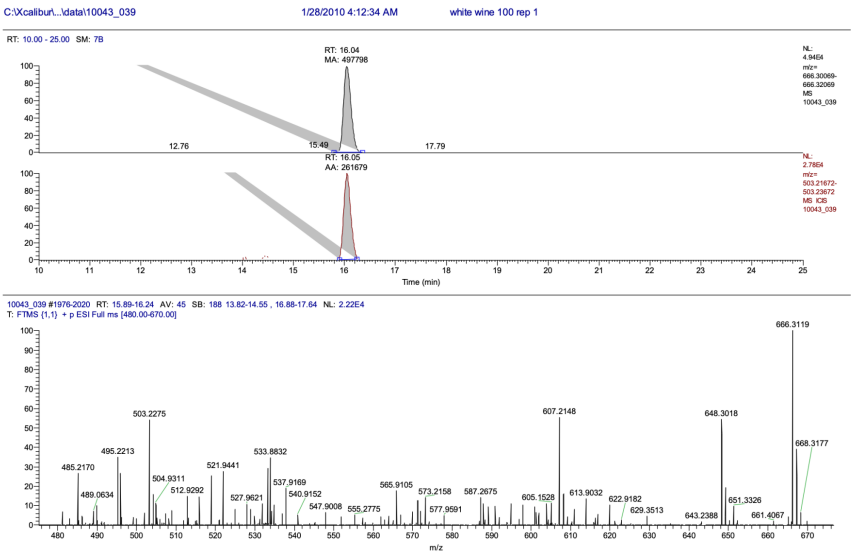
2.1.5.2. Quantitation

Natamycin determination for samples with an estimated concentration of greater than 5 µg/l. Pipette 2 ml of wine into five 2 ml Eppendorf centrifuge vials and add 0 µl, 5 µl, 10 µl, 20 µl and 50 µl of natamycin working solution 1 (2.1.3.3.2) into the vials respectively. This is equivalent to 0 µg/l, 25 µg/l, 50 µg/l, 100 µg/l and 250 µg/l natamycin added. Shake the vials for one minute and then centrifuge for 10 min at 14000 rpm. Filter an aliquot through 0.2 µm PTFE into an amber 2 ml vial. Analyse by LC-HR/MS (section 6) and estimate the concentration of natamycin in the sample (section 7).

2.1.6. ANALYSIS

NOTE: When starting measurements baseline stability and response linearity of the detector should be examined, together with verification of the detection limit. Maintain the same operating conditions throughout the measurement of all samples and calibration standards. Identify the natamycin peaks on the basis of the retention time and their accurate mass channel, and measure the peak areas. Inject each of the solutions as prepared onto the LC column. Measure the peak area of the natamycin peak in each of the quantification and confirmation channels. An example of a typical chromatogram is given in Figure 1.

Figure 1. Typical LC-HR/MS chromatogram and mass spectrum for natamycin spiked into white wine at the equivalent of 50 µg/l in the sample.



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Plot the peak area for the main quantification channel against the concentration of natamycin added in micrograms per litre (µg/l). Determine the slope, intercept point and correlation coefficient of the regression line. The calibration curve shall be rectilinear and the correlation coefficient shall be 0.99 or better.

2.1.7. EXPRESSION OF RESULTS

2.1.7.1. Calculation of analyte level

The natamycin concentration in the sample in micrograms per litre (µg/l) is calclulated using the following formula:

C = b/a

where C = concentration of natamycin in the wine (µg/l), a = slope of the regression line, b = y-intercept point of the regression line

2.1.8. CONFIRMATION

The presence of natamycin in the samples shall be confirmed by applying the following criteria:

The presence of a peak in both accurate mass channels m/z 666.31069 and m/z 503.22672 at the same retention time. Calculate the ratio of the peak area for the main quantification mass channel relative to the peak area of the confirmation channel. The criterion is that the ratios agree to ± 25% of those obtained from the standard addition calibration standards.

2.1.9. METHOD PERFORMANCE DATA

2.1.9.1. Linearity

The method is linear over the calibration range of 1 to 2640 µg/l in solvent, white wine and red wine matrices (figures 2, 3 and 4).

Figure 2. Ten point calibration graph of natamycin spiked into solvent in the range from 1 to 2600 µg/l.

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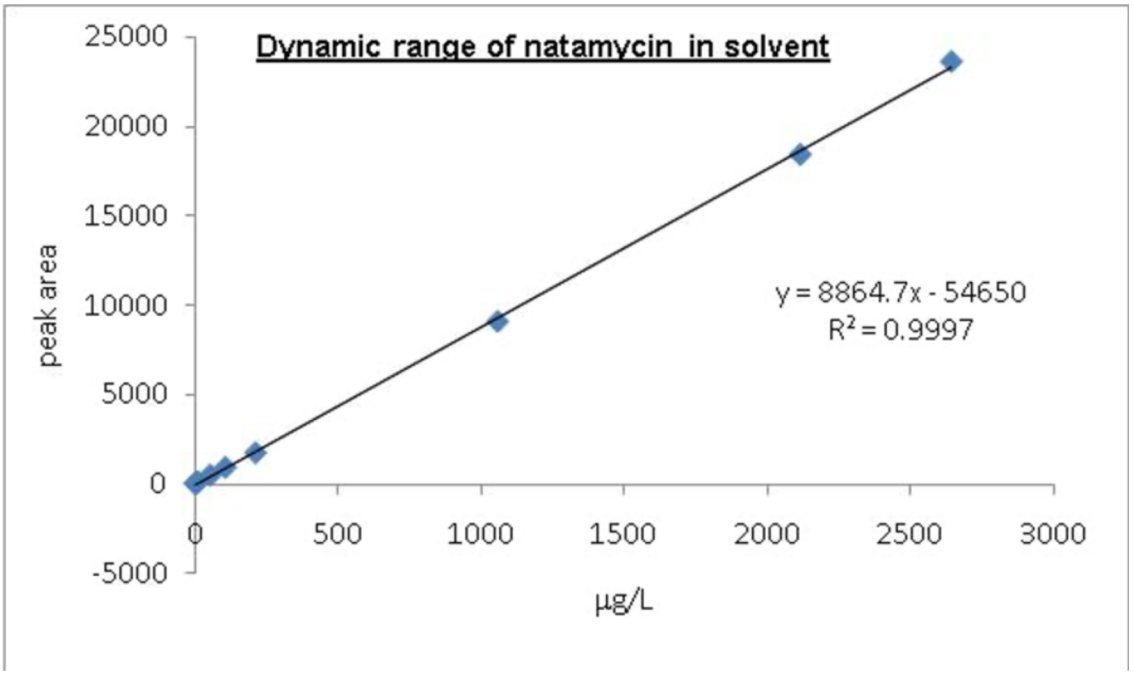


Table 1. Solvent calibration residuals.

Natamycin (µg/l)	Predicted conc. (µg/l)	Residuals	Standard Residuals
0	6.4	-6.4	-0.4
1.056	6.8	-5.7	-0.3
5.28	10.9	-5.6	-0.3
10.56	16.8	-6.3	-0.4
52.8	58.9	-6.1	-0.3
105.6	108.3	-2.7	-0.2

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211.2	200.1	11.1	0.6
1056	1029.8	26.2	1.5
2112	2084.8	27.2	1.6
2640	2671.8	-31.8	-1.8

Figure 3. Ten point calibration graph of natamycin spiked into white wine in the range from 1 to 2600 µg/l.

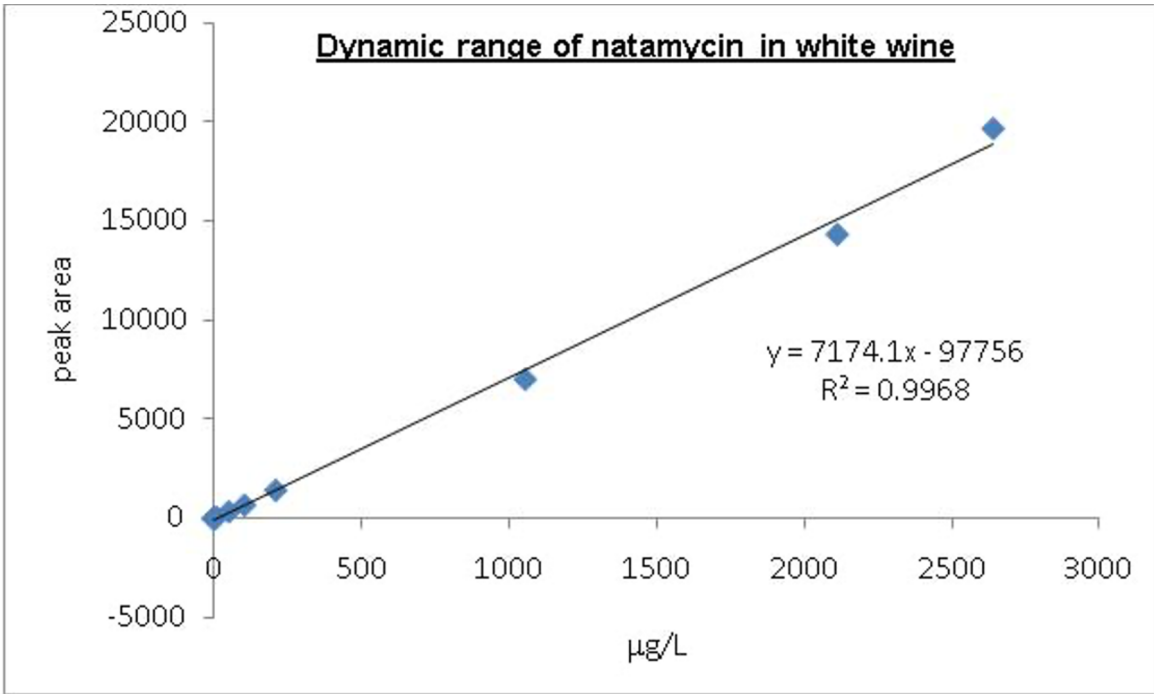


Table 2 White wine matrix calibration residuals.

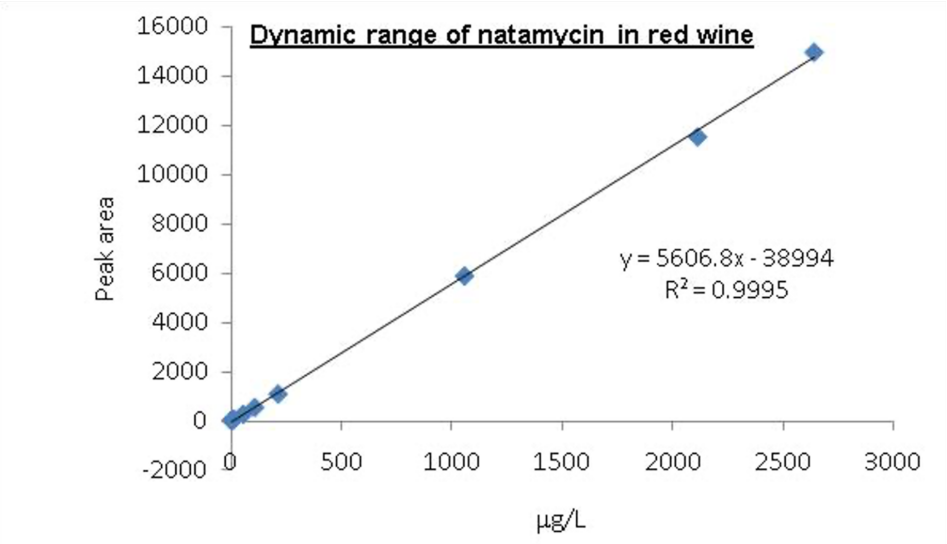
Natamycin (µg/l)	Predicted conc. (µg/l)	Residuals	Standard Residuals

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0	15.5	-15.5	-0.3
1.056	15.6	-14.6	-0.3
5.28	18.8	-13.5	-0.2
10.56	23.9	-13.3	-0.2
52.8	63.6	-10.8	-0.2
105.6	109.3	-3.7	-0.1
211.2	212.8	-1.6	0.0
1056	989.0	67.0	1.2
2112	2003.2	108.8	2.0
2640	2742.7	-102.7	-1.8

Figure 4. Ten point Calibration graph of natamycin spiked into red wine in the range from 1 to 2600 µg/l.



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Table 3. Red wine matrix calibration residuals.

Natamycin (µg/l)	Predicted conc. (µg/l)	Residuals	Standard Residuals
0	7.2	-7.2	-0.3
1.056	8.2	-7.1	-0.3
5.28	10.9	-5.7	-0.3
10.56	16.8	-6.2	-0.3
52.8	52.1	0.7	0.0
105.6	102.1	3.5	0.2
211.2	199.8	11.4	0.5
1056	1055.2	0.8	0.0
2112	2063.7	48.3	2.3
2640	2678.4	-38.4	-1.8

2.1.9.2. Accuracy and Precision

The method was assessed for repeatability at the interventiont limit of 5 µg/l and at 200 µg/l in solvent, white wine and red wine matrices (tables 4, 5 and 6). The accuracy was assessed by spiking a known amount at two different levels into white and red wine. The analysis was then performed by a second analyst without the knowledge of the spiked natamycin concentration. The results are shown in table 7.

Table 4. Repeatability of natamycin spiked into solvent (methanol:water:acetic acid, 50:47:3 v/v) at two concentrations; 5 and 200 µg/l.

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	Conc. Natamycin ug/l	Recovery (%)
Solvent std at 5 ng/ml rep 1	5.3	99.7
Solvent std at 5 ng/ml rep 2	5.4	101.8
Solvent std at 5 ng/ml rep 3	5.8	108.6
Solvent std at 5 ng/ml rep 4	5.7	108.2
Solvent std at 5 ng/ml rep 5	5.8	109.0
Solvent std at 5 ng/ml rep 6	5.9	112.2
Solvent std at 5 ng/ml rep 7	5.7	108.4
Solvent std at 5 ng/ml rep 8	6.4	120.2
Average	5.8	108.5
Std deviation	0.3	6.2
RSD (%)	5.7	5.7
Solvent std at 200 ng/ml rep 1	238.3	112.9
Solvent std at 200 ng/ml rep 2	237.1	112.4
Solvent std at 200 ng/ml rep 3	231.5	109.7

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Solvent std at 200 ng/ml rep 4	228.0	108.1
Solvent std at 200 ng/ml rep 5	244.0	115.7
Solvent std at 200 ng/ml rep 6	220.7	104.6
Solvent std at 200 ng/ml rep 7	229.4	108.7
Solvent std at 200 ng/ml rep 8	251.7	119.3
Average	235.1	111.4
Std deviation	9.8	4.7
RSD (%)	4.2	4.2

Table 5. Repeatability of natamycin spiked into white wine at two concentrations; 5 and 200 µg/l.

	Conc. Natamycin ug/l	Recovery (%)
White wine 5.3 ng/ml rep 1	5.3	99.1
White wine 5.3 ng/ml rep 2	4.4	82.8
White wine 5.3 ng/ml rep 3	5.1	96.0
White wine 5.3 ng/ml rep 4	4.9	92.5

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White wine 5.3 ng/ml rep 5	4.6	86.4
White wine 5.3 ng/ml rep 6	5.1	96.4
White wine 5.3 ng/ml rep 7	4.8	90.9
White wine 5.3 ng/ml rep 8	4.9	92.2
Average	4.9	92.0
Std deviation	0.3	5.4
RSD (%)	5.9	5.9
White wine 211 ng/ml rep 1	217.6	103.1
White wine 211 ng/ml rep 2	223.3	105.8
White wine 211 ng/ml rep 3	213.0	101.0
White wine 211 ng/ml rep 4	216.8	102.7
White wine 211 ng/ml rep 5	211.4	100.2
White wine 211 ng/ml rep 6	208.6	98.9
White wine 211 ng/ml rep 7	204.2	96.8
White wine 211 ng/ml rep 8	214.4	101.6
Average	213.7	101.3
Std deviation	5.8	2.8

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RSD (%)	2.7	2.7

Table 6. Repeatability of natamycin spiked into red wine at two concentrations; 5 and 200 µg/l.

	Conc. Natamycin ug/l	Recovery (%)
Red wine 5.3 ng/ml rep 1	5.3	99.7
Red wine 5.3 ng/ml rep 2	5.0	93.8
Red wine 5.3 ng/ml rep 3	3.8	72.5
Red wine 5.3 ng/ml rep 4	5.1	96.5
Red wine 5.3 ng/ml rep 5	5.0	95.0
Red wine 5.3 ng/ml rep 6	5.5	103.5
Red wine 5.3 ng/ml rep 7	4.3	80.9
Red wine 5.3 ng/ml rep 8	4.8	90.7
Average	4.9	91.6
Std deviation	0.5	10.2
RSD (%)	11.1	11.1
Red wine 211 ng/ml rep 1	183.9	87.1

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Red wine 211 ng/ml rep 2	178.4	84.5
Red wine 211 ng/ml rep 3	181.1	85.8
Red wine 211 ng/ml rep 4	197.5	93.6
Red wine 211 ng/ml rep 5	178.2	84.5
Red wine 211 ng/ml rep 6	184.2	87.3
Red wine 211 ng/ml rep 7	181.2	85.9
Red wine 211 ng/ml rep 8	171.3	81.2
Average	182.0	86.2
Std deviation	7.5	3.6
RSD (%)	4.1	4.1

Table 7. Accuracy of natamycin spiked into white and red wine at two concentrations; 125 and 220 µg/l.

	Theoretical concentration (µg/l)	**Obtained concentration (µg/l)**	**Accuracy (%)**	**Z Score**
White wine A rep 1	125	135	108	0.50
White wine A rep 2	125	142	114	0.85

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White wine A rep 3	125	138	110	0.65
White wine B rep 1	220	230	105	0.28
White wine B rep 2	220	230	105	0.28
White wine B rep 3	220	239	109	0.54
Red wine A rep 1	220	213	97	-0.20
Red wine A rep 2	220	234	106	0.40
Red wine A rep 3	220	223	101	0.09
Red wine B rep 1	125	129	103	0.20
Red wine B rep 2	125	129	103	0.20
Red wine B rep 3	125	120	96	-0.25

Calculations

Z scores calculated as:

= (obtained concentration - theoretical concentration)/ target standard deviation

Where:

Target standard deviation = 0.16 x spiked concentration

i.e according to Horwitz

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2.2. Détermination of natamycin (pimaricin) in wine by HPLC/DAD

2.2.1. Scope

This method describes an analytical procedure for the determination of natamycin (pimaricin) in wine by HPLC. The level of natamycin is expressed in micrograms per litre (µg/l) of wine.

The described method has been laboratory validated taking into account the influence of the matrix wine (e.g. white wine or red wine).

2.2.2. Principle

Non-sparkling wine samples are directly injected into the HPLC system. Sparkling wine samples are degased first by filtration or by using an ultrasonic bath. The analyte is separated from the matrix on a C8-column. The fraction window with the analyte is automatically transferred to a C18-column for further separation. Natamycin is detected at 304 nm and 319 nm. Additionally the DAD spectrum is used for identification. Quantification is done with reference to external standards.

2.2.3. Reagents and Material

2.2.3.1. Reagents

2.2.3.1.1. Water, deionised

2.2.3.1.2. Methanol, HPLC grade (CAS no. 67-56-1).

2.2.3.1.3. Formic acid, p. a. (CAS no. 64-18-6).

2.2.3.1.4. Acetic acid, p. a. (CAS no. 64-19-7).

2.2.3.1.5. Hydrochloric acid, p. a., 0,1 N (CAS no. 7647-01-0).

2.2.3.1.6. Matrix wine, natamycin not detectable

2.2.3.1.7. Natamycin, > 95 % (CAS no. 7681-93-8).

The purity is verified by photometric measurement at 291 nm, 304 nm and 319 nm of a natamycin solution in hydrochloric acid, 0,1 N against a blank of hydrochloric acid, 0,1 N:

Reference data according to the literature	291 nm	304 nm	319 nm
Extinction (1 Gew.% Natamycin, 1 cm cell)	758	1173	1070

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Alternative:

After dilution (e. g. dilution factor 20) the stock solution (2.1.3.3.1.) can also be used for the photometric measurement, e. g. pipette 1,0 ml stock solution into a 20 ml volumetric flask and fill up to the mark with hydrochloric acid, 0,1 N. Measure against a blank with the same composition of solvents as the diluted stock solution.

2.2.3.2. Preparation of the mobile phase

2.2.3.2.1. Solutions for the mobile phase:

2.2.3.2.1.1. 5 ml acetic acid added to 2 l methanol

2.2.3.2.1.2. 5 ml acetic acid added to 2 l deionised water

2.2.3.2.2. Eluent 1: methanol-acetic acid / deionised water-acetic acid (65 / 35)

2.2.3.2.3. Eluent 2: methanol-acetic acid / deionised water-acetic acid (80 / 20)

2.2.3.3. Preparation of the stock and standard solutions

All solutions have a limited stability and have to be stored dark and cold in a refrigerator. The stock solution (2.1.3.3.1.1) has a shelf life up to several weeks but the concentration has to be checked shortly before usage (e.g. see alternative method, 2.2.3.1.7.). Dilution I (2.2.3.3.1.2) and II (2.2.3.3.1.3) and the standard solutions (2.2.3.3.2) have to be prepared daily.

2.2.3.3.1. Preparation of the stock solution and dilutions

2.2.3.3.1.1. Stock solution (approximately 100 mg/l)

Weight in about 5 mg natamycin (3.1.7) and transfer with methanol into a 50 ml volumetric flask. Add 0,5 ml formic acid, make sure that all the natamycin is dissolved, temperate at 20 °C and make up to the mark with methanol.

2.2.3.3.1.2. Dilution I (approximately 5 mg/l)

Pipette 2,5 ml of the stock solution (2.1.3.3.1.1) into a 50 ml volumetric flask and make up to the mark with deionised water.

2.2.3.3.1.3. Dilution II (approximately 1 mg/l)

Pipette 4 ml of Dilution I (2.2.3.3.1.2) into a 20 ml volumetric flask and make up to the mark with the matrix wine (2.2.3.1.6).

2.2.3.3.2. Preparation of the standard solutions

For the standard solutions dilute Dilution II (2.2.3.3.1.3) to the desired concentrations with the matrix wine (2.2.3.1.6), e. g. 50 µl into a 10 ml volumetric flask equals 5 µg/l:

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Volumetric flask	10 ml	10 ml	10 ml	10 ml	10 ml	10 ml	10 ml
Volume of Dilution II (µl)	50	100	200	400	500	1000	3700
Amount of natamycin (µg/l)	5	10	20	40	50	100	370

2.2.4. Apparatus

Usual laboratory equipment, in particular the following:

2.2.4.1. HPLC-DAD apparatus with a 6 port HPLC valve and two isocratic pumps or a gradient pump to enable fractionation

2.2.4.2. HPLC-column RP-8

2.2.4.3. HPLC-column RP-18

2.2.4.4. Photometer

2.2.5. Sampling

Non-sparkling wine samples are directly injected into the HPLC system. Sparkling wine samples are first degased by filtration or by using an ultrasonic bath. If samples need to be stored the storage conditions should be cold and dark.

2.2.6. Procedure

2.2.6.1. Operating conditions of HPLC

The following columns and parameters have been found to be suitable:

Column 1:	C 8-column (e.g. Select B 125*4mm/5µm endcapped, Merck)
Mobile phase:	Eluent 1 (2.2.3.2.2) at room temperature
Flow rate:	1 ml/min
Column 2:	C 18-column (e.g. Lichrospher 125*4mm/5µm, Merck)
Mobile phase:	Eluent 2 (2.2.3.2.3) at 30°C

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Flow rate:	1 ml/min
Injection volume:	500 µl
UV-detection:	304 nm and 319 nm
Fraction window:	The position of the fraction window has to be checked prior to the following analysis (fig. 1). The range of the fraction window has to be set at 0,5 min. before and after the desired peak elutes from the C 8-column.

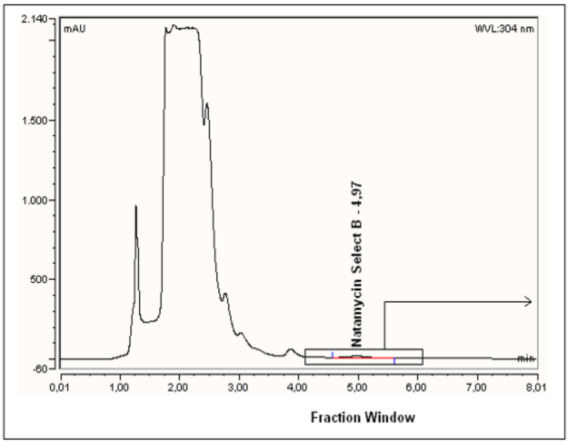


Fig. 1 Column 1

Fraction window

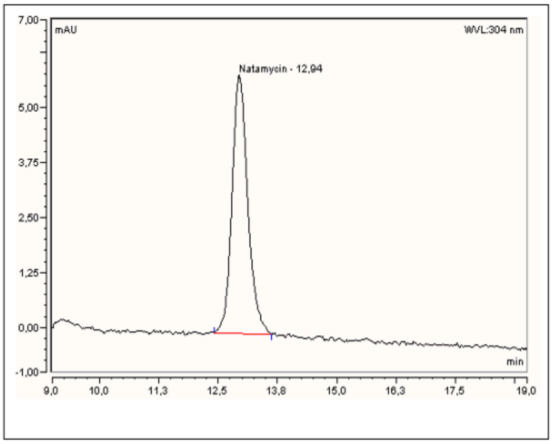


Fig. 2 Column 2

White wine spiked with natamycin (50 µg/l)

2.2.6.2. Identification/ Confirmation

Identification of peaks is done by the comparison of retention times between standards and samples for both measured wavelengths 304 nm and 319 nm. Using the chromatographic system and parameters of 2.2.6.1 the retention time for natamycin is approximately 12,9 min (fig. 2).

The DAD spectrum is used for further confirmation of positive findings (fig. 3 and fig. 4).

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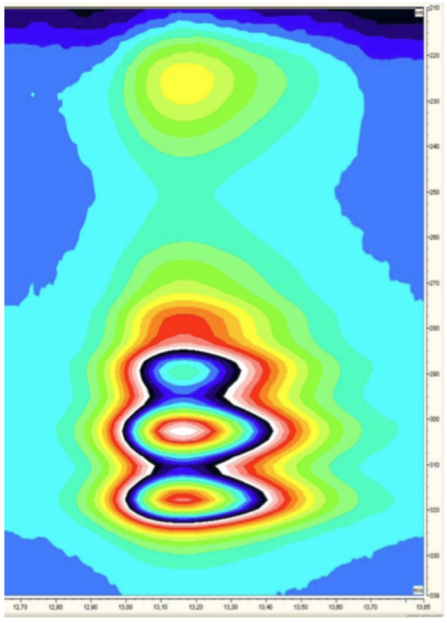


Fig. 3 3D-DAD spectra of natamycin

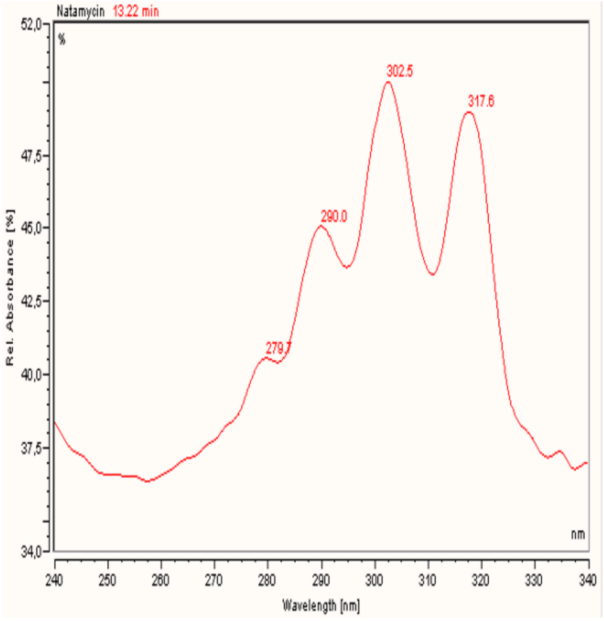


Fig. 4 DAD spectra of natamycin

2.2.7. Calculation and expression of results

A calibration curve of the standard solutions (2.2.3.3.2) is prepared using the chromatograms measured at 304 nm. The quantification of natamycin is performed following the external calibration method. A linear calibration curve is generated by comparison of the peak areas and the relevant concentrations. The correlation coefficient should be at least 0,99.

The expression of the results is µg/l.

2.2.8. Method performance data

Detection limit, Quantification limit

The detection limit and quantification limit were determined according to DIN 32645 (direct determination: multiple measurement of a blank matrix sample, n=10, and a calibration curve that covers the total working range).

Detection limit: 2,5 µg/l

Quantification limit: 8,5 µg/l

Linearity

The linearity in a wine matrix is confirmed in the calibration range of 5 µg/l to 100

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µg/l (fig. 5).

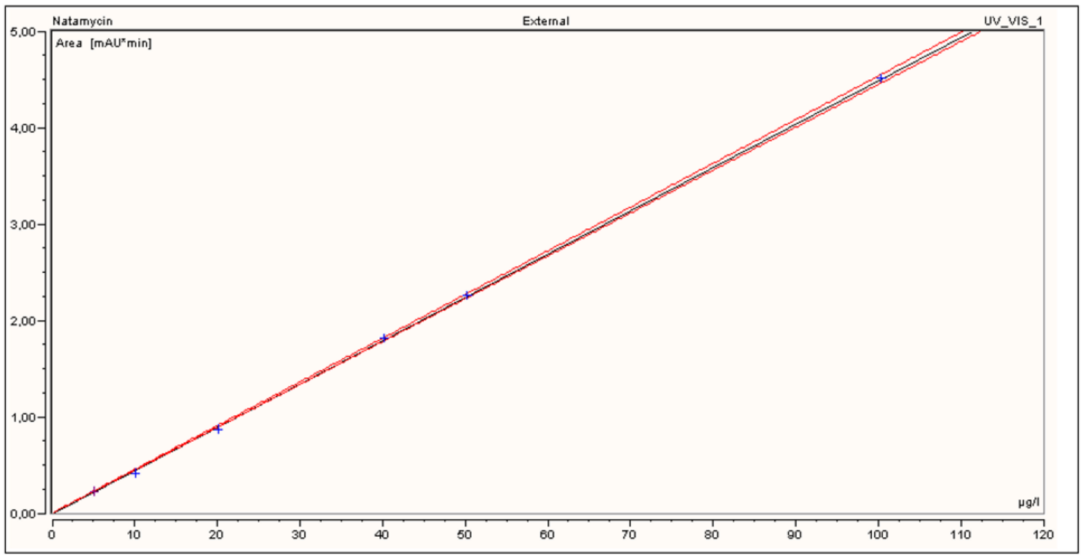


Fig. 5 Six point calibration graph of natamycin spiked into white wine matrix in the range from 5 to 100 µg/l, R^2=0,9999

2.2.9. Trueness and Precision

Trueness and repeatability were assessed by spiking a known amount of natamycin into white, rosé and red wine and measuring each of these samples five times. The results are shown in table 1.

Matrix	Natamycin content in matrix (µg/l)	Spiked natamycin content (µg/l)	Measured natamycin content (µg/l)	Recovery rate (%)	Z-Score
White wine	n. d.	5.02	5.04	100.4	0.0
			4.70	93.6	-0.2
			5.12	102.0	0.1
			5.29	105.4	0.2
			4.97	99.0	0.0
		Average	5.02	100.1	
		Std dev.	0.22		
		RSD (%)	4.3		
		Repeatability r	0.85		

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Rosé wine	n. d.	5.02	4.79	95.4	-0.1
			4.83	96.2	-0.1
			4.76	94.8	-0.1
			4.79	95.4	-0.1
			4.73	94.2	-0.2
			4.78	95.2	
			0.04		
			0.78		
			0.15		
			4.61	91.8	-0.2
			4.65	92.6	-0.2
			4.89	97.4	-0.1
			4.67	93.0	-0.2
			4.34	86.5	-0.4
			4.63	92.3	
			0.20		
			4.2		
			0.77		
			19.73	93.1	-0.2
			20.66	97.5	-0.1
			21.16	99.8	0.0
			19.73	93.1	-0.2
			19.58	92.4	-0.3
			20.17	95.2	
			0.70		
			3.5		
			2.7		
			51.84	97.4	-0.1
			51.91	97.6	-0.1
			51.42	96.7	-0.1
			50.12	94.2	-0.2
			50.62	95.2	-0.2
			51.18	96.2	
			0.78		
			1.5		
			3.1		

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Table 1 Accuracy of natamycin spiked into white. rosé and red wine; n.d. "not detected". detection limit 2.5 µg/l

Calculations (Table 1):

Repeatability r = Std dev. * t 4;0.95 * 2 1/2

Z score = (measured amount-spiked amount)/ target standard deviation *

* according to Horwitz

target standard deviation = 1/100 * spiked amount * $2^{(1-0.5 \log spiked\ amount}$

References

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